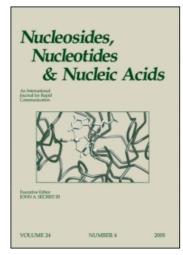
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Potential Antiviral Agents. Part II. Synthesis and Antiviral Evaluation of Pyrazinones Substituted With Acyclic Chains

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POTENTIAL ANTIVIRAL AGENTS. PART II. SYNTHESIS AND ANTIVIRAL EVALUATION OF PYRAZINONES SUBSTITUTED WITH ACYCLIC CHAINS.

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Abstract: The synthesis of a series of 4'-substituted hydroxybutyl pyrazine analogues of the anti-herpes compound, acyclovir, is described. The compounds were characterized with ¹H and ¹³C nmr, mass and IR spectroscopy. Antiviral (HSV-1, CMV, Cox B4, HIV-1) properties of these compounds were examined. None of these compounds were active against these viruses.

Since the discovery of 9-[2-(hydroxyethoxy)methyl]guanine (acyclovir)¹, a selective antiherpes virus agent, considerable interest has been focused on the synthesis of novel acyclic analogues of nucleosides.² As a result of this research, a number of hydroxyalkylated derivatives of guanine have been identified as potential antiviral drugs.³ Recently, certain 6-substituted acyclic pyrimidine nucleosides related to acyclovir, such as 1-[2-hydroxyethoxy)methyl]-6-(phenylthio)thymine (HEPT) have been shown to be selective inhibitors of human immunodeficiency virus in various human lymphocytes.⁴ With this last family of compounds, it has been found that an intact hydroxyl is not necessary for its antiviral activity.⁵

After having synthesised several pyrazinic analogues of acyclovir and evaluated their antiviral potential⁶, several modifications of these molecules were studied - in particular, the transformation of the 4' position of the hydroxybutyl chain of

hydroxyalkylated pyrazinones by azidation (6a,b,c and 7a,b,c) and by glycosylation (9a,b,c and 11a,b,c) [Scheme]. The synthesis and antiviral activites (HSV-1, CMV, Cox B4, vaccine, HIV-1) in cell cultures of this series of alkyl pyrazinones will be reported herein.

Results and Discussion

The 3-substituted pyrazinones (1a,b) were synthesised according to Jones method⁷ and 1c using a modified method described in our preceding paper.⁸

Starting from 1a-c, the synthesis of compounds 6a-c, 7a-c and 10a-c and 11a-c is outlined in Scheme I. Compounds 2a-c and 3a-c were prepared by reacting sodium hydride with the appropriate pyrazinone followed by the addition of bromobutylacetate using the conditions previously described⁶. Deacetylation of compounds 2a-c and 3a-c with sodium methoxide occurred readily; 4a-c and 5a-c were recovered in quasi quantitative yields.

Compounds 6a-c and 7a-c were prepared by the initial mesylation of alkyl pyrazinones in the presence of mesyl chloride in anhydrous pyridine at 0°C and were used without further purification. Azidation was carried out in DMF at 100°C in the presence of NaN₃. After purification, the obtained yields are comparable to literature values (38-65% overall yield).

Compounds 8a-c were obtained using the method of Koenigs-Knorr⁹ which reacts a glycosidic derivative with an alcohol in stoechiometric quantities in the presence of a catalyst (SnCl₄, TiCl₄,AgTf etc....). Thus, 4a, 4b and 4c were reacted with 2,3,4,6-tetra-O-acetyl-α-D-glucopyranosyl bromide in the presence of silver trifluoromethane sulfonate and s-collidine in dichloromethane protected from light. The s-collidine was added either at the beginning of the reaction according to Vazquez¹² or at the end of the reaction to neutralize the medium; the addition mode did not affect the final yield. After purification, compounds 8a-c were obtained in approximately 20% yields. Compounds 8a-c are formed according to the classic anchimeric assistance mechanism of β-glycosylation. In all cases, there was significant acetate transfer from the O-2 to the 4'-hydroxyl of the glycosyl group. This acetate transfer is not uncommon since it has often been reported to compete with glycosylation reactions.

i) NaH, Br(CH₂)₄OAc, DMF. ii) MeONa, McOH. iii) MsCl, pyridine then NaN₃, DMF. iv) 2,3,4,6-tetra-O-acetyl- α -**D**-glucopyranosyl bromide, AgTf, CH₂Cl₂. v)1-O-acetyl-2,3,5-tri-O-benzoyl- β -**D**-ribofura - nose, SnCl₄, CH₃CN.

Scheme

Thus even as the desired products were successfully synthesised, the yields of these glycosylation reactions remained the limiting step. Others have also experienced difficulties in glycosylating certain hydroxyl groups¹⁴, in particular, the primary 5'-hydroxyl group of nucleosides using standard conditions for the formation of glycosidic bonds^{15,16}. Krepinsky *et al*¹⁶ have postuled that the π -electron systems of the heterocyclic base or the heteroaromatic bonds interact with the unreactive hydroxyl group. They confirmed through semiempirical and *ab initio* molecular modeling

calculations of several nucleoside model compounds that intramolecular hydrogen bonding between the ribosyl O5'—H······O2 of the base is a stabilizing factor in their cytosine model compound. The existence of such intramolecular hydrogen bonding in nucleosides is apparently well documented.¹⁷ In the present case, it is likely that the alcohol function is hydrogen bonded with the 2-keto group.

The removal of the acetyl groups from 8 to give the unprotected compounds 9 proceeded readily with sodium methoxide in nearly quantative yields. 18

Compounds 10 a,b,c were obtained using the method of Hannessian.¹¹ Thus 4a, 4b and 4c were reacted with 1-O-acetyl-2,3,5-tri-O-benzoyl-β-D-ribofuranose in the presence of Tin(IV) chloride in acetonitrile. After purification, compounds 10 a-c were obtained with 49, 45 and 42% overall yields respectively. In this case a significative acetylation of aglycone (40%) was also observed.

The removal of benzoyl groups from 10 to give the unprotected compounds 11 proceeded readily with sodium methoxide in nearly quantitative yields. 19

Biological Evaluation

Most of the prepared compounds were tested for their *in vitro* inhibitory effects on the replication of a number of DNA viruses (herpes simplex virus type 1, human cytomegalovirus, vaccinia virus) and RNA viruses (Coxsackie virus B4, HIV-1).

As previously reported, no antiviral activity was observed with compounds substituted with an hydroxybutyl group. Modification of the 4' position of the hydroxylalkyl group by azidation (6a-c, 7a-c) or glycosylation (9a-c, 10a-c) did not improve their activity.

When evaluated in two anti-human immunodeficiency virus (anti-HIV 1) assays, none of the tested compounds showed an antiviral effect at a concentration less than the minimum concentration causing a detectable alteration of MT-4 and CEM host cell viability.

Experimental

Thin-layer chromatography (TLC) was performed on silica gel Kieselgel 60PF₂₅₄ (Merck) plates and visualized in several ways: by an ultraviolet light source at 254 nm

and/or 365 nm, by spraying with sulfuric acid (6N) and heating to 200°C, by vaporizing with a fluoresceine solution followed by an aqueous solution of hydrogen peroxide in acetic acid (for compounds containing Br) or by a combination of two or more of these techniques. Silica gel (Merck Kieselgel 60, 15-40 µm) was used for flash chromatography. Solvents were distilled from appropriate drying agents. Solutions were concentrated at 1 Torr pressure in a rotary evaporator. ¹H and ¹³C NMR spectra were recorded at 300 (75, 13C) MHz with a Bruker AM-300 spectrometer or at 200 (50, ¹³C) MHz with a Bruker Ac-200 spectrometer. Chemical shifts (δ) are expressed in ppm with Me₄Si as internal standard (δ =0). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, qt = quintet, m = multiplet and br = broad), coupling constants in hertz (Hz) and assignment. Electronic-impact mass spectra (EI) were recorded with a Shimadzu QP1000 mass spectrometer at the Laboratoire Départemental d'Analyses de Limoges. Chemical-impact mass spectra (CI) were recorded with a Kratos MS580 mass spectrometer; Fast Atom Bombardment (FAB) spectra were recorded on a R10-10 Nermag spectrometer. Both CI and FAB were recorded at the Laboratoire de Chimie Organique Structurale of the Université Pierre et Marie Curie (Paris VI). Melting points (°C) were determined with a Kofler block and are uncorrected. Elemental analyses were carried out by Microanalytical Service of the Université Pierre et Marie Curie (Paris VI). Rotatory dispersions were measured with a Jasco (DIP-370) polarimeter in a 1 dm quartz cell at 22°C. Infra-red spectra (KBr disk or film) were measured on a Perkin Elmer 1310 grating spectrophotometer and are reported in wave numbers (cm⁻¹). UV spectra were recorded with a Hewlett Packard 8454A diode array spectrophotometer. corresponding to the maximum absorbances, λ_{max} , are expressed in nanometers and the molar absorptivity coefficients, ε in mol⁻¹, l. cm⁻¹, are expressed as their log values.

Compounds 4a-c and 5a-c were synthesised according to ref 6.

N^{1} -(4-azidobutyl)-2-pyrazinone (6a)

To a solution of N¹-(4-hydroxybutyl) 2-pyrazinone 4a (230 mg, 1.4 mmol) in a minimum of pyridine (4 mL, anhydrous 99.8%) at 0°C was added 4 equivalents of mesyl chloride (0.42 mL). After 1 hour, the solution was quenched with the addition of a

saturated NaHCO₃ solution / H₂O (10 mL, 1:1, v/v) and extracted with CHCl₃ (3 x 15 mL). The organic layer was dried (MgSO₄), filtered and the solvent removed. The resultant product (258 mg, 78% crude yield) was dissolved in DMF (3 mL). This solution was immersed in an oil bath at 100°C and sodium azide (340 mg, 7 mmol) was added. After 1 hour, the product was extracted with 3 x 20 mL chloroform and the extracts were evaporated. The crude product (141 mg) was purified using preparative tlc (CH₂Cl₂ / EtOH, v/v, 95:5). Pure N¹-(4-azidobutyl) 2-pyrazinone was recovered in an overall yield of 45% (118 mg). $R_f = 0.5$, (CH₂Cl₂/MeOH, 95/5, v/v). IR: 3070-3050 cm⁻¹ (CH (aromatic)), 2950-2850 cm⁻¹ (CH), 2180 cm⁻¹ (N₃), 1650 cm⁻¹ (C=O), 1580 cm⁻¹ (C=C (aromatic)). ¹H (CDCl₃): δ 8.14 (1H, d, J=1.1, H₃), 7.08 (1H d, J=4.4-1.1, H₅), 7.32 (1H, d, J=4.4, H₆), N-alkyl: 3.92 (2H t, J=7.1, H₁), 1.84 (2H, m, H₂), 1.64 (2H, m, H₃·), 3.35 (2H, t, J= 6.5, H₄). ¹³C (CDCl₃): δ 156.4 (C₂), 149.8 (C₃), 128.3 (C₅), 123.7 (C₆) N-alkyl 48.8 (C₁), 25.9 (C₂), 26.0 (C₃·), 50.8 (C₄·). Anal. calcd. for C₈H₁₁ON₅ requires C 49.73, H 5.74, N 36.25, found C 49.57, H 5.75, N 35.99. MS (DCI/NH₃) m/z 194 (MH¹).

Compounds 6b and 6c were prepared in a manner similar to that described for 6a starting from 4b and 4c respectively.

N^{I} -(4-azidobutyl)-3-methyl 2-pyrazinone (6b)

Yield 65% (168 mg). $R_f = 0.49$, (CHCl₃/MeOH, 95/5, v/v). IR: 3080-3050 cm⁻¹ (CH (aromatic)), 2950-2820 cm⁻¹ (CH), 2160 cm⁻¹ (N₃), 1640 cm⁻¹ (C=O), 1580 cm⁻¹ (C=C (aromatic)). H (CDCl₃): δ 6.97 (1H, dd, J=4.4-0.6, H₅), 7.18, (1H, d, J=4.5, H₆), 2.44, (3H, d, J=0.6, H_{methyl}), N-alkyl: 3.90 (2H t, J=7.0, H₁), 1.84 (2H, m, H₂), 1.64 (2H, m, H₃), 3.34 (2H, t, J=6.5, H₄). H₃C (CDCl₃): δ 158.6 (C₂), 156.5 (C₃), 126.8 (C₅), 122.6 (C₆), 20.8 (C_{methyl}), N-alkyl: 48.9 (C₁), 25.9 (C₂), 25.9 (C₃), 50.8 (C₄). Anal. calcd. for C₉H₁₀ON₅ C 52.16, H 6.32, N 33.79, found C 52.16, H 6.34, N 33.63. MS (DCI/NH₃) m z 208 (MH⁺).

N^{1} -(4-azidobutyl)-3-decyl 2-pyrazinone (6c)

Yield 61% (163 mg). $R_f = 0.45$ (CH₂Cl₂). IR: 3100 cm⁻¹ (CH (aromatic)), 2950-2850 cm⁻¹ (CH), 2100 cm⁻¹ (N₃), 1655 cm⁻¹ (C=O), 1600 cm⁻¹ (C=C (aromatic)). ¹H (CDCl₃)

δ 6.96 (1H, br.d, J = 4.4, H₅), 7.22 (1H, d, J = 4.4, H₆), decyl chain: 0.86 (3H, t, J = 6.4, H₁₀), 1.25 (14H, br. s, H₃₋₉), 1.74 (2H, m, H₂), 2.80 (2H, t, J = 7.5, H₁), N-alkyl 3.90 (2H t, J = 7.1, H₁), 1.85 (2H, m, H₂), 1.64 (2H, m, H₃), 3.34 (2H, t, J = 6.6, H₄). ¹³C (CDCl₃) δ 161.6 (C₂), 155.9 (C₃), 126.4 (C₅), 122.5 (C₆), decyl chain 14.0 (C₁₀), 22.7 (C₉), 29.3 (2C, C₇₋₈), 29.5 (4C, C₃₋₆), 31.9 (C₂), 32.5 (C₁), N-alkyl. 48.9 (C₁), 26.5 (C₂), 26.0 (C₃), 50.8 (C₄). Anal calcd for C₁₈H₃₁ON₅ calculated C 64.83, H 9.37, N 21.00, found C 64.95, H 9.51, N 20.91. MS (DCI/NH₃) m/z 334 (MH⁺).

O-(4-azidobutyl)-2-pyrazinone (7a)

The residue obtained from reaction of mesyl chloride with **5a** (288 mg, 1.7 mmol) was reacted with sodium azide. After one hour and work up, the product (125 mg, 75%) was extracted and purified using preparative tlc with CH₂Cl₂ as eluent. Pure 7a was recovered in 42% yield (70 mg). $R_f = 0.65$, (CHCl₃/EtOH, 95/5, v/v). IR: 2990-2900 cm⁻¹ (CH), 2170 cm⁻¹ (N₃), 1210 cm⁻¹ (C-O-C (aryl alkyl)). ¹H (CDCl₃): δ 8.19 (1H, m, H₃), 8.10 (1H, m, H₅), 8.10 (1H, m, H₆), *N-alkyl*: 4.36 (2H, t, *J*=6.1, H₁), 1.85 (2H, m, H₂), 1.85 (2H, m, H₃), 3.35 (2H, t, *J*=6.5, H₄). ¹³C (CDCl₃): δ 160.4 (C₂), 140.6 (C₃), 138.3 (C₅), 135.8 (C₆) *N-alkyl* 65.5 (C₁), 26.0 (C₂), 25.6 (C₃), 51.1 (C₄). Anal calcd for C₈H₁₁ON₅ calculated C 49.73, H 5.74, N 36.25, found C 50.40, H 5.90, N 37.27. MS (DCI/NH₃) m/z 194 (MH⁺).

Compounds 7b and 7c were prepared in a manner similar to that described for 7a starting from 5b and 5c (0.06 mmol) respectively.

O-(4-azidobutyl)-3-methyl-2-pyrazinone (7b)

Yield 42% (94 mg). $R_f = 0.67$, (CHCl₃/MeOH, 9/1, v/v). IR: 3080 cm⁻¹ (CH (aromatic)), 2950-2870 cm⁻¹ (CH (aliphatic)), 2100 cm⁻¹ (N₃), 1550 cm⁻¹ (C=C). 1170 cm⁻¹ (COC). ¹H (CDCl₃): δ 7.89 (1H, dd, J=2.9-0.6, H₅), 7.98 (1H, d, J=2.9. H₆), 2.44 (3H, br.s, H_{methyl}), O-alkyl: 4.36 (2H t, J=6.1, H₁), 1.85 (2H, m, H₂), 1.85 (2H, m, H₃), 3.35 (2H, t, J=6.5, H₄). ¹³C (CDCl₃): δ 158.5 (C₂), 144.8 (C₃), 138.1 (C₅), 133.5 (C₆), 19.2 (C_{methyl}), O-alkyl: 65.4 (C₁), 26.1 (C₂), 25.6 (C₃), 51.1 (C₄). Anal calcd for C₉H₁₀ON₅ C 52.16, H 6.32, N 33.79, found C 52.22, H 6.41, N 33.74. MS (DCI/NH₃) m/z 208 (MH⁺).

O-(4-azidobutyl)-3-decyl-2-pyrazinone (7c)

Yield 38%. $R_f = 0.58$, (CHCl₃), $R_{f \text{ (mesyl)}} = 0.44$, (CH₂Cl₂/MeOH, 95/5, v/v). IR: 3080 cm⁻¹ (CH (aromatic)), 2950-2870 cm⁻¹ (CH (aliphatic)), 2100 cm⁻¹ (N₃), 1550 cm⁻¹ (C=C), 1170 cm⁻¹ (COC). ¹H (CDCl₃) δ 6.96 (1H, br.d, J = 4.4, H₅), 7.22 (1H, d, J = 4.4, H₆), decyl chain: 0.86 (3H, t, J = 6.4, H₁₀), 1.25 (14H, br. s, H₃₋₉), 1.74 (2H, m, H₂), 2.80 (2H, t, J = 7.5, H₁), N-alkyl 4.36 (2H t, J = 6.0, H₁·), 1.84 (2H, m, H₂·), 1.84 (2H, m, H₃·), 3.37 (2H, t, J = 6.6, H₄·). ¹³C (CDCl₃) δ 158.2 (C₂), 148.4 (C₃), 138.0 (C₅), 135.4 (C₆), decyl chain 14.1 (C₁₀), 22.7 (C₉), 29.3 (2C, C₇₋₈), 29.5 (4C, C₃₋₆), 31.9 (C₂), 32.5 (C₁), N-alkyl. 65.4 (C₁·), 26.2 (C₂·), 25.7 (C₃·), 54.1 (C₄·). Anal calcd for C₁₈H₃₁ON₅ C 64.83, H 9.37, N 21.00, found C 64.66, H 9.48, N 21.10. MS (DCI/NH₃) $m \in \mathbb{Z}$ 334 (MH⁺).

N^{1} -(4-(2'',3'',4'',6''-tetra-O-acetyl- β -D-glucopyranosyloxy)-butyl)-2-pyrazinone (8a)

To a dry system of N¹-(4-hydroxybutyl) 2-pyrazinone (80 mg, 0.36 mmol) in freshly distilled CH2Cl2 (6 mL) under Ar and protected from light were added molecular sieves (4Å), 1-bromo 2,3,4,6-tetra-O-acetyl-β-D-glucopyranose (275 mg, 1.4 eq.) and AgOTf (170 mg, 1.4 eq.). The system was vigorously stirred until completion of reaction as monitered by tlc and neutralized with the addition of s-collidine. The mixture was allowed to stir for an additional 30 minutes and then filtered through celite. The product was purified using preparative silica gel plates with an elution system of toluene/acetone/pet. ether in a 4:5:1 ratio and was recovered in 22% yield (40 mg). $R_f =$ 0.53, (toluene/acetone/pet. ether: 3:6:1, v/v/v). $[\alpha]_D +19.1^\circ$ (0.3, CHCl₃). ¹H (CDCl₃) δ 8.08 (1H, d, J = 1.5, H₃), 7.04 (1H, dd, J = 1.5 - 4.3, H₅), 7.25 (1H, d, J = 4.3, H₆) Nalkyl 3.68 (2H, t, J = 7.2, C_1H), 1.60 (2H, m, C_2H), 1.77 (2H, m, C_3H), 4.05 (2H, t, J =6.2, C₄·H), glucopyramose 4.44 (1H, d, J=8.0, H_{1"}), 4.90 (1H, dd, J=8.0-9.5, H_{2"}), 5.12 (1H, t, J = 9.5, $H_{3''}$), 5.01 (1H, t, J = 9.8, $H_{4''}$), 3.63 (1H, m, $H_{5''}$), 4.34 (1H, dd, J = 12.3-4.8, $H_{6''a}$), 4.09 (1H, dd, J = 12.0 - 2.4, $H_{6''b}$), 2.05-1.95 (12H, 4s, $4H_{Acetyl}$). MS $(DCI/NH_3) m/z 499 (MH^+).$

Compounds 8b and 8c were prepared in a manner similar to that described for 8a starting from 4b and 4c (0.5 and 0.3 mmol) respectively.

N^{l} -(4-(2",3",4",6"-tetra-O-acetyl- β -D-glucopyranosyloxy)-butyl)-3-methyl-2-pyrazinone (8b)

Yield 20%. $R_f = 0.38$, (toluene/acetone/pet. ether, 4:5:1, v/v/v). $[\alpha]_D + 16.8^\circ$ (0.33, CHCl₃). 1H (CDCl₃) δ 7.02 (1H, d, J = 4.4, H₅), 7.20 (1H, d, J = 4.4, H₆), 2.47 (3H, s, H_{methyl}), N-alkyl 3.90 (2H, t, J = 6.2, H₁), 1.83 (2H, m, H₂), 1.67 (2H, m, H₃), 3.90 (2H, t, H₄), glucopyranose 4.50 (1H, d, J = 8.1, H_{1"}), 4.98 (1H, dd, J = 8.2-9.3, H_{2"}), 5.21 (1H, t, J = 9.4, H_{3"}), 5.09 (1H, t, J = 9.5, H_{4"}), 3.70 (1H, m, H_{5"}), 4.26 (1H, dd, J = 12.1-4.6, H_{6"a}), 4.14 (1H, .dd, J = 12.1-2.8, H_{6"b}), 2.08-1.98 (12H, 4s, 4H_{Acetyl}). MS (DCI/NH₃) m/z 513 (MH⁺).

N^{l} -(4-(2'',3'',4'',6''-tetra-O-acetyl- β -D-glucopyranosyl)-butoxy)-3-decyl-2-pyrazinone (8c)

Yield 15%. $R_f = 0.40$, (toluene/acetone/pet. ether, (6:2:2) x2, v/v/v). $[\alpha]_D + 47.7^\circ$ (0.40, CHCl₃). 1H (CDCl₃) δ 6.91 (1H, d, J = 4.1, H₅), 7.14 (1H, d, J = 4.1, H₆), decyl chain: 0.85 (3H, t, J = 5.4, H₁₀), 1.26 (14H, br. s, H_{3.9}), 1.68 (2H, m, H₂), 2.76 (2H, t, J = 7.7, H₁), N-alkyl 3.87 (2H t, J = 7.8, H₁), 1.83 (2H, m, H₂), 1.58 (2H, m, H₃), 3.91 (2H, t, J = 5.3, H₄), glucopyranose 4.50 (1H, d, J = 8.3, H₁), 4.99 (1H, dd, J = 9.7 - 8.3, H₂), 5.21 (1H, t, J = 9.7, H₃), 5.08 (1H, t, J = 9.6, H₄), 3.70 (1H, m, H₅), 4.27 (1H, dd, J = 11.7 - 5.0, H₆, 4.15 (1H, dd, J = 11.7 - 3.2, H₆, 2.04-1.97 (12H, 4s, 4H_{Acetyl}). MS (DCI/NH₃) m/z 639 (MH⁺).

(4-(β-D-glucopyranosyl) butoxy)-2-pyrazinone (9a)

The deacetylation of the parent compound (40 mg, 0.080 mmol) was carried out in the presence of 0.5 eq of sodium methoxide (1M solution in methanol.). When tlc showed reaction completion, the solution was then neutralized by addition of Amberlite IRN 77 H⁺ resin (Aldrich). When neutral pH was reached, the solution was quickly filtered and the resin thoroughly rinsed with methanol. 9a was obtained in 85% yield (23 mg, 0.068 mmol). $R_f = 0.52$, (CHCl₃/EtOH, 1:2, v/v). [α]_D +35.4° (0.30, CHCl₃). ¹H (D₂O/CD₃OD) δ 8.04 (1H, d, J = 1.4, H₃), 7.61 (1H, dd, J = 1.5 - 4.3, H₅), 7.40 (1H, d, J = 4.2, H₆) *N-alkyl* 3.59 (2H, t, J = 7.0, C₁H), 1.88 (2H, m, C₂H), 1.76 (2H, m, C₃H), 3.94 (2H, t, J = 6.3, C₄H), *glucopyranose* 4.27 (1H, d, J = 7.7, H₁°), 3.97 (1H, dd, J = 7.7, H₁°), 3.97 (1H, dd, J = 7.7, H₂°), 3.97 (1H, dd, J = 7.7, H₃°), 3.97 (1H, dd, J = 7.7, H₄°), 3.97 (1H, dd, J = 7.7, H₁°), 3.97 (1H, dd, J = 7.7, H₂°), 3.97 (1H, dd, J = 7.7, H₃°), 3.97 (1H, dd, J = 7.7, H₄°), 3.97 (1H, dd, J = 7.7, H₁°), 3.97 (1H, dd, J = 7.7, H₁

7.7-7.5, $H_{2"}$), 4.03 (1H, t, J = 7.4, $H_{3"}$), 4.02 (1H, t, J = 7.6, $H_{4"}$), 3.66 (1H, m, $H_{5"}$), 3.86 (1H, dd, J = 11.7-1.7, $H_{6"a}$), 3.81 (1H, dd, J = 11.7-2.0, $H_{6"b}$). MS (DCI/NH₃) m/z 331 (MH⁺).

Compounds 9b and 9c were prepared in a manner similar to that described for 9a starting from 8b and 8c (0.06 mmol) respectively.

$(4-(\beta-D-glucopyranosyloxy)-butyl)-3-methyl-2-pyrazinone (9b)$

Yield 87%. $R_f = 0.45$, (CHCl₃/EtOH, 1:1, v/v) x 2. $[\alpha]_D + 48.2^\circ$ (0.40, CHCl₃). ¹H (D₂O/CD₃OD) δ 7.23 (1H, dd, J = 4.4-1.8, H₅), 7.34 (1H, d, J = 4.3, H₆), 2.44 (3H. s. H_{methyl}), N-alkyl 3.62 (2H, t, J=7.0, H₁), 1.87 (2H, m, H₂), 1.68 (2H, m, H₃), 3.94 (2H, m, H₄), glucopyranose 4.26 (1H, d, J = 7.8, H_{1"}), 3.96 (1H, m, H_{2"}), 4.00 (1H, t, J = 7.7, H_{3"}), 3.98 (1H, t, J = 7.3, H_{4"}), 3.68 (1H, m, H_{5"}), 3.87 (1H, dd, J = 11.8-2.4, H_{6"a}), 3.79 (1H, dd, J = 11.9-2.5, H_{6"b}). MS (DCI/NH₃) m/z 345 (MH⁺).

$(4-(\beta-D-glucopyranosyloxy)-butyl)-3-decyl-2-pyrazinone (9c)$

Yield 92%. $R_f = 0.48$, (CHCl₃/EtOH, 1:1, v/v). $[\alpha]_D + 39.1^\circ$ (0.25, CH₃OH). ¹H (CDCl₃/CD₃OD) δ 7.51 (1H, d, J = 4.3, H₅), 7.23 (1H, d, J = 4.3, H₆), decyl chain: 0.85 (3H, t, J = 5.5, H₁₀), 1.31 (14H, br. s, H_{3.9}), 1.71 (2H, m, H₂), 2.82 (2H, t, J = 7.7, H₁), N-alkyl 3.63 (2H t, J = 7.0, H₁), 1.87 (2H, m, H₂), 1.74 (2H, m, H₃), 3.96 (2H, m, H₄), glucopyranose 4.27 (1H, d, J = 7.5, H₁₀), 3.98 (1H, m, H₂₀), 4.05 (1H, t, J = 7.6, H₃₀), 4.04 (1H, t, J = 7.3, H₄₀), 3.69 (1H, m, H₅₀), 3.83 (1H, dd, J = 11.8 - 2.4, H_{60a}), 3.79 (1H, dd, J = 11.9 - 1.8, H_{60b}). MS (DCI/NH₃) m/z 472 (MH⁺).

N^{l} -(4-(2", 3", 5"-tri-O-benzoyl- β -D-furanosyloxy)- butyl)-2-pyrazinone (10a)

To a dry system of N^1 -(4-hydroxybutyl) 2-pyrazinone **4a** (60 mg, 0.36 mmol) in freshly distilled acetonitrile at O°C were added 1-O-acetyl-2,3,5-tri-O-benzoyl- β -D-ribofuranose (300 mg, 1 eq.) and tin (IV) chloride (2 eq.). After completion of reaction as monitered by tlc, the solution was quenched with the addition of a saturated NaHCO₃ solution / H₂O (20 mL, 1:1, v/v) and extracted with CH₂Cl₂ (3 x 20 mL). The organic layer was dried (MgSO₄), filtered and the solvent removed. The product was purified using flash chromatography with an elution gradient of ethyl acetate / methanol and was

recovered in 49% yield (180 mg). $R_f = 0.41$ ($CH_2Cl_2/MeOH$, 95:5, v/v). 1H ($CDCl_3$) δ 8.08 (1H, d, J=1.0, H₃), 7.10 (1H, br. d, J=4.6 Hz, H₅), 7.31 (1H, d, J=4.5 H₆), N-alkyl 3.87 (2H t, J=7.3, H₁), 1.62 (2H, m, H₂), 1.80 (2H, m, H₃), 3.49 (1H, dt, J=9.7-6.1, H_{4'a}), 3.83 (1H, dt, J=9.7-6.1, H_{4'b}), ribose 5.23 (1H, s, H_{1''}), 5.66 (1H, br. d, J=4.9, H_{2''}), 5.84 (1H, dd, J=6.5-4.9, H_{3''}), 4.70 (1H, m, H_{4''}), 4.52 (1H, dd, J=12.8-6.5, H_{5''a}), 4.72 (1H, dd, J=12.8-4.1 Hz, H_{5''b}), benzoyl groups 7.89, 8.02, 8.06 (6H, dd, J=8.5-1.5, H_{2.6}), 7.36-7.51 (9H, m, H_{3,4.5}). ^{13}C ($CDCl_3$) δ 149.6 (C_2), 149.5 (C_3), 123.4 (C_5), 123.8 (C_6), N-alkyl 49.0 (C_1), 25.5 (C_2), 26.4 (C_3), 67.8 (C_4), ribose 105.6 (C_1 "), 75.5 (C_2 "), 72.4 (C_3 "), 79.0 (C_4 "), 64.8 (C_5 "), benzoyl groups 128.9,129.1, 129.2 (C_1), 128.4, 128.5, 128.6 (C_3 .5), 129.7 (C_2 .6), 133.2, 133.4, 133.5 (C_4), 165.3, 165.4, 166.1 (C_7). MS (DCI/NH_3) m/z 613 (MH^+).

Compounds 10b and 10c were prepared in a manner similar to that described for 10a starting from 4b and 4c (0.65 and 0.50 mmol) respectively.

N^{I} -(4-(2'',3'',5''-tri-O-benzoyl- β -D-furanosyloxy)- butyl)-3-methyl-2-pyrazinone (10b)

Yield 45% (190 mg). R_f 0.53 (EtOAc). ¹H (CDCl₃) δ 7.03 (1H,br.d, *J*=4.5, H₅), 7.18 (1H, d, *J*=4.5, H₆), C-alkyl 2.49 (3H, s, CH₃), *N-alkyl* 3.87 (2H t, *J*= 7.3, H₁), 1.60 (2H, m, H₂), 1.80 (2H, m, H₃), 3.49 (1H, dt, *J*=9.7-6.1, H_{4*a}), 3.83 (1H, dt, *J*=9.7-6.1, H_{4*b}), *ribose* 5.23 (1H, s, H_{1*}), 5.66 (1H, br. d, *J*=4.9, H_{2*}), 5.84 (1H, dd, *J*=6.5-4.9, H_{3*}), 4.70 (1H, m, H_{4*}), 4.52 (1H, dd, *J*=12.8-6.6, H_{5*a}), 4.72 (1H, dd, *J*=12.8-4.1, H_{5*b}), *benzoyl groups* 7.89, 8.02, 8.06 (6H, dd, *J*=8.5-1.5, H_{2.6}), 7.36-7.51 (9H, m, H_{3,4,5}). ¹³C (CDCl₃) δ 149.5 (C₂), 149.6 (C₃), 123.4 (C₅), 123.8 (C₆), C-alkyl 20.6 (CH₃), *N-alkyl* 49.0 (C_{1*}), 25.5 (C_{2*}), 26.4 (C_{3*}), 67.8 (C_{4*}), *ribose* 105.6 (C_{1*}), 75.5 (C_{2*}), 72.4 (C_{3*}), 79.0 (C_{4*}), 64.8 (C_{5*}), *benzoyl groups* 128.9,129.1, 129.2 (C₁), 128.4, 128.5, 128.6 (C_{3,5}), 129.7 (C_{2.6}), 133.2, 133.4, 133.5 (C₄), 165.3, 165.4, 166.1 (C₇). MS (DCI/NH₃) m/z 627 MH⁺.

 N^{l} -(4-(2",3",5"-tri-O-benzoyl- β -D-furanosyloxy)- butyl)-3-decyl-2-pyrazinone (10c) Yield 42% (158 mg). R_f 0.53 (CH₂Cl₂/EtOH, 95/5, v/v). ¹H (CDCl₃) δ 7.03 (1H,br.d, J=4.5, H₅), 7.18 (1H, d, J=4.5, H₆), C-alkyl 2.83 (2H,br.t, J=7.8, CH₂), 1.7 (2H, m,

CH₂), 1.3 (14H, br.s, CH₂) 0.88 (3H, br.t, *J*=7.4, CH₃), *N*-alkyl 3.88 (2H t, *J*= 7.3, H₁), 1.70 (4H, m, H_{2′,3′}), 3.49 (1H, dt, *J*=9.7-6.1, H_{4′a}), 3.83 (1H, dt, *J*=9.7-6.1, H_{4′a}), *ribose* 5.23 (1H, s, H_{1′′}), 5.66 (1H, br. d, *J*=4.9, H_{2′′}), 5.84 (1H, dd, *J*=6.5-4.9, H_{3′′}), 4.70 (1H. m, H_{4′′}), 4.52 (1H, dd, *J*=12.8-6.6, H_{5′′a}), 4.72 (1H, dd, *J*=12.8-4.1 , H_{5′′b}), *benzoyl groups* 7.89, 8.00, 8.06 (6H, dd, *J*=8.5-1.5, H_{2.6}), 7.36-7.51 (9H, m, H_{3.4.5}). ¹³C (CDCl₃) 8 155.8 (C₂), 161.3 (C₃), 121.3 (C₅), 127.3 (C₆), C-alkyl 33.2 (C₁), 31.9 (C₂), 29.3, 29.4, 29.6, 26.7, 22.8 (C_{3·9}), 14.1 (C₁₀), *N*-alkyl 49.3 (C_{1′}), 25.5 (C₂), 26.4 (C_{3′}), 67.8 (C_{4′}), *ribose* 105.6 (C_{1′′}), 75.5 (C_{2′′}), 72.5 (C_{3′′}), 79.0 (C_{4′′}), 64.8 (C_{5′′}), *benzoyl groups* 128.9,129.1, 129.2 (C₁), 128.4, 128.5, 128.6 (C_{3.5}), 129.7 (C_{2.6}), 133.2, 133.4, 133.5 (C₄), 165.3, 165.4, 166.1 (C₇). MS (DCI/NH₃) m/z 753 MH⁴.

Compoud 11a, 11b and 11c were prepared in a manner similar to that described for 9a starting from 10a, 10b and 10c (0.39, 0.20 and 0.50 mmol) respectively.

N^{l} (4- β -D-ribofuranosyloxy)-butyl)-2-pyrazinone (11a)

Yield 89% (105 mg). R_f 0.28 (CHCl₃/MeOH, 80/20, v/v). ¹H (CDCl₃) δ 8.04 (1H, d, J=0.9, H₃), 7.6 (1H, dd, J=0.9-4.5 Hz, H₅), 7.4 (1H, d, J=4.5, H₆), N-alkyl 4.0 (2H t, J=7.6, H₁), 1.6 (2H, m, H₂), 1.84 (2H, m, H₃), 3.43 (1H, dt, J=6.3-6.2, H_{4'a}), 3.80 (1H, dt, J=6.3-6.2, H_{4'b}), ribose 4.84 (1H, s, H_{1"}), 3.87 (1H, d, J=4.9, H_{2"}), 4.05 (1H, dd, J=6.9-4.9, H_{3"}), 3.94 (1H, td, J=6.9-3.6, H_{4"}), 3.54 (1H, dd, J=12.1-6.9, H_{5"a}), 3.74 (1H, dd, J=12.1-3.6 Hz, H_{5"b}). ¹³C (CDCl₃) δ 157.2 (C₂), 148.2 (C₃), 124.5 (C₅), 130.6 (C₆), N-alkyl 49.5 (C_{1'}), 25.7 (C_{2'}), 26.6 (C_{3'}), 67.2 (C_{4'}), ribose 107.8 (C_{1"}), 75.3 (C_{2"}), 71.6 (C_{3"}), 83.8 (C_{4"}), 63.9 (C_{5"}). MS (DCI/NH₃) m/z 301 (MH⁺).

N^{I} -(4- β -D-ribofuranosyloxy)-butyl)-3-methyl-2-pyrazinone (11b)

Yield 86% (67 mg). R_f 0.35 (CH₂Cl₂/EtOH, 80/20, v/v). 1 H (CDCl₃) δ 7.6 (1H, d, J=4.5 Hz, H_5), 7.4 (1H, d, J=4.5, H_6), C-alkyl 2.30 (3H, s, CH₃), N-alkyl 4.1 (2H t, J= 7.6, H_1), 1.65 (2H, m, H_2), 1.88 (2H, m, H_3), 3.43 (1H, dt, J=6.5-6.5, H_4 ₁₂), 3.80 (1H, dt, J=6.5-6.5, H_4 ₁₃), ribose 4.82 (1H, s, H_1 ¹³), 3.88 (1H, d, J=4.9, H_2 ¹³), 4.05 (1H, dd, J=6.7-4.9, H_3 ¹³), 3.94 (1H, td, J=6.7-3.6, H_4 ¹³), 3.54 (1H, dd, J=12.2-6.7, H_5 ¹³), 3.74 (1H, dd, J=12.2-3.6 Hz, H_5 ¹³). 13 C (CDCl₃) δ 157.2 (C₂), 158.2 (C₃), 124.5 (C₅), 130.6 (C₆), C-10.5 (CDCl₃)

alkyl 20.5 (CH₃), *N-alkyl* 49.5 (C₁), 25.7 (C₂), 26.6 (C₃), 67.2 (C₄), *ribose* 107.8 (C_{1°}), 75.3 (C_{2°}), 71.6 (C_{3°}), 83.8 (C_{4°}), 63.9 (C_{5°}). MS (DCI/NH₃) m/z 315 (MH⁺).

N^{l} -(4- β -D-ribofuranosyloxy)-butyl)-3-decyl-2-pyrazinone (11c)

Yield 79% (79 mg). R_f 0.53 (CH₂Cl₂/EtOH, 80/20, v/v). 1 H (CDCl₃) δ 7.25 (1H,d, J=4.5, H_5), 7.43 (1H, d, J=4.5, H_6), C-alkyl 2.75 (2H, t, J=7.8, CH₂), 1.7 (2H, m, CH₂),1.3 (14H, br.s, CH₂), 0.89 (3H, t, J=7.0, CH₃), N-alkyl 3.97 (2H t, J=7.6, H_1), 1.7 (2H, m, H_2), 1.8 (2H, m, H_3), 3.42 (1H, dt, J=6.3-6.4, H_4 ₂), 3.8 (1H, dt, J=6.3-6.4, H_4 ₃), ribose 4.87 (1H, s, H_1 ²), 3.87 (1H, d, J=4.9, H_2 ²), 4.05 (1H, dd, J=7.0-4.9, H_3 ²), 3.92 (1H, td, J=3.6-7 0, H_4 ²), 3.54 (1H, dd, J=12.2-7.0, H_5 ²₃), 3.72 (1H, dd, J=12.2-3.6, H_5 ²₅. 13 C (CDCl₃) δ 156.6 (C₂), 160.5 (C₃), 122.9 (C₅), 128.6 (C₆), C-alkyl 33.4 (C₁), 32.1 (C₂), 29.7, 29.6, 29.5, 29.4, 26.8 (C₃.9), 13.5 (C₁₀), N-alkyl 49.6 (C₁), 25.7 (C₂), 26.7 (C₃), 67.2 (C₄), ribose 107.8 (C₁²), 71.6 (C₂²), 63.9 (C₃²), 83.8 (C₄²), 63.9 (C₅²). MS (DCI/NH₃) m/z 441 MH⁺.

Biological Methods

The antiviral assays carried out on human embryonic fibroblasts (Cell line MRC 5) infected with coxsackie virus B4 (Cox B4), Herpes simplex virus type 1 (HSV-1), Human Cytomegalovirus (CMV) and Vaccinia virus were described previousely. ¹⁹ The antiviral activity is expressed as the IC₅₀ concentration necessary to reduce viral cytopathicity by 50%.

Cytotoxicity (MTT assay)

Cell viability was evaluated by measuring the activity of mitochondrial dehydrogenase using the MTT assay.²⁰

Anti-HIV 1 assays

The anti HIV 1 activity was tested on CEM-SS and MT₄ cells infected respectively with HIV 1 LAI and HIV 1 IIIB following protocols described previously.²¹

References

- ¹ G. B. Elion, P. A. Furman, J. A. Fyle, P. Miranda, L. Beauchamp, H. J. Schaeffer, *Proc. Natl. Acad. Sci. USA*, 74, 5716 (1977).
- ² For reviews, see: C. K. Chu & S. J. Cutler, *J. Heterocycl. Chem.*, **23**, 289, (1986); R.J. Remy & J. A. Secrist, *Nucleosides, Nucleotides*, **4**, 411 (1985).
- (a) A. Larsson, S. Alenius, N.-G. Johansson, B. Oberg, Antiviral Res., 3, 77 (1983); (b)
 D. Sutton, M.R. Boyd, Antimicrob. Agents Chemother., 37, 642 (1993); (c) D.L. Earnshaw, T.H. Bacon, S.J. Darlison, K. Edmonds, M. Perkins, R.A. Hodge, Antimicrob. Agents Chemother., 36, 2747 (1992); (d). C.K. Chu, S.J.J. Cutler, J. Heterocyclic Chem., 1986, 23, 289; (e) K.K. Ogilvie, N. Nguyen-Ba, M.F. Gillen, B.K. Radatus, U.O. Cheriyan, H.R. Hanna, K.O. Smith, K.S. Galloway, Can. J. Chem., 62, 241 (1984); (f) A. Genevois, J. -C. Florent, C. Monneret, D.S. Grierson, Tetrahedron Lett., 31(34), 4879 (1990).
- ⁴ H. Tanaka, M. Baba, M. Ubasawa, H; Takashima, K. Sekiya, I. Nitta, S. Shigeta, R.T. Walker, E. DeClercq, T. Miyasaka, *J. Med. Chem.*, **34**, 1394 (1991) and refs. therein.
- ⁵ M. Baba, E. De Clercq, H. Tanaka, M. Ubasawa, H. Takashima, K. Sekiy, I. Nitta, R.T. Walker, S Mori, M. Ito, S. Shigeta, T. Miyasaka, *Proc. Natl. Proc. Sci. USA*, **88**, 2356 (1991).
- ⁶ J. Davis, R. Benhaddou, R. Granet, P. Krausz, M. De Monte, A.-M. Aubertin, Nucleosides & Nucleotides (in press).
- ⁷ R. Jones, J. Am. Chem. Soc., 71, 78 (1949).
- ⁸A. Benjahad, R.Granet, P.Krausz, C. Bosgiraud, S. Delebassée, *Nucleosides & Nucleotides*, **15**, 1849 (1996).
- ⁹ W. Koenigs, E. Knorr, Ber. Dtsch. Chem. Ges., 34, 957 (1901).
- ¹⁰ H. Paulsen, Angew. Chem. Int. Ed. Engl., 21, 155 (1982).
- ¹¹ (a) S. Hanessian, J. Banoub, J. Carbohydr. Res., 53, 13 (1977); (b) S. Hanessian, J. Banoub, in Methods in Carbohydrate Chemistry, Academic Press, eds., 8, 243 (1980);

- (c) T. Ogawa, K. Beppu, S. Nakabayashi, *Carbohydr. Res.*, C6, 93 (1981); (d) R.R. Schmidt, *Angew. Chem. Int. Ed. Engl.*, 25, 212 (1986); (e) P. Fügedi, P.J. Garegg, H. Lönn, T. Norberg, *Carbohydr. Res.*, C13, 177 (1987).
- ¹² M. Trujillo, E.Q. Morales, J.T. Vazquez, J. Org. Chem., **59**, 6637 (1994).
- (a) D.M. Whitfield, S.P. Douglas, T.-H. Tang, I.G. Csizmadia, H.Y.S. Pang, F.L. Moolten, J.J. Krepinsky, Can. J. Chem., 72, 2225, (1994); (b) T.-H. Tang, D.M. Whitfield, S.P. Douglas, J.J. Krepinsky, I.G. Csizmadia, Can. J. Chem., 72, 1803, (1994); (c) P.J. Garegg, P. Konradsson, I. Kvarnström, T. Norberg, S.C. Svensson, B. Wigilius, Acta Chem. Scand.B, 39, 569 (1985); (d) T. Zielger, P. Kovac, C.P. Glaudemans, Ann. Chem., 613 (1190); (e) N.I. Uvarona, G.I. Oshitok, G.B. Elyakov, Carbohydr. Res., 37, 79 (1973); (f) A.Y. Khorlin, V.A. Nesmeyanov, S.E. Zurabyan, Carbohydr. Res., 43, 69 (1975); (g) R.U. Lemieux, Chem. Can., 16, 14 (1964); (h) J. Banoub, D.R. Bundle, Can. J. Chem., 57, 2091 (1979); (i) G. Wulff, G. Röhle, Angew. Chem. Int. Ed. Engl., 13, 157 (1974).
- 14 (a) H. Paulsen, Angew. Chem. Int. Ed. Engl., 21, 155 (1982), Angew. Chem. Int. Ed. Engl., 29, 823 (1990); (b) A.H. Haines, Adv. Carbohydr. Chem. Biochem., 33, 11 (1976).
- (a) D.M. Whitfield, S.P. Douglas, J.J. Krepinsky, *Tetrahedron Lett.*, 33 (45), 6795 (1992); (b) S.P. Douglas, D.M. Whitfield, J.J. Krepinsky, Fuji '90 Post-Symposium: Recent Progress of Synthetic Methods in Carbohydrates and their Applications to Synthetic Chemistry. Susono City, Shizuoka, Japan, 1990. (c) T.-H. Tang, D.M. Whitfield, S.P. Douglas, J.J. Krepinsky, I.G. Csizmadia, *Can. J. Chem.*, 70, 2434, (1992); (d) D.M. Whitfield, S.P. Douglas, T.-H. Tang, I.G. Csizmadia, H.Y.S. Pang, F.L. Moolten, J.J. Krepinsky, *Can. J. Chem.*, 72, 2225, (1994); (e) T.-H. Tang, D.M. Whitfield, S.P. Douglas, J.J. Krepinsky, I.G. Csizmadia, *Can. J. Chem.*, 72, 1803, (1994).
- ¹⁶ (a) F.W. Lichtenthaler, Y. Sanemitsu, T. Nohara, Angew. Chem. Int. Ed. Engl., 17, 772 (1978); (b) N.D. Chkanikov, M.N. Preobrazhenskaya, Nucleosides Nucleotides., 8, 391 (1981); (c) N.B. Hanna, R.K. Robins, G.R. Revenkar, Carbohydr. Res., 165, 267 (1987).

¹⁷ (a) G.I. Birnbaum, D. Shugar, in Topics in Molecular and Structural Biology, S. Neidle, W. Fuller, eds., MacMillian Press, New York, USA, 9, pp. 1-77, 1987 and references therein; D. Plochocka, A. Rabczenko, D.B. Davies, J. Chem. Soc. Perkin Trans. I, 2, 82 (1981); (b) G.I. Birnbaum, F.E. Hruska, W.P. Niemczura, J. Am. Chem. Soc., 102, 5586 (1980).

- ¹⁸ G. Zemplen, A. Kunz, Chem. Ber., 56, 1705 (1923).
- O. Génu-Dellac, G. Gosselin, A.-M. Aubertin, G. Obert, A. Kirn, J.-L. Imbach. Antiviral Chemistry & Chemotherapy, 1991, 2(2), 83.
- ²⁰ T. Mossman, J. Immunol. Methods, 65, 55 (1983).
- ²¹ C. Moog, A. Wick, P. LeBer, A. Kirn, A.-M. Aubertin, *Antiviral Res.*, **24**, 275 (1994).

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